

# International Journal of Engineering Research and Science & Technology

ISSN : 2319-5991  
Vol. 3, No. 2  
May 2014



Review Article

## ESCHERICHIA COLI AS A MODEL ORGANISM

**Muhammad Kamran Taj<sup>1</sup>, Zohra Samreen<sup>2</sup>, Ji Xiu Ling<sup>1</sup>, Imran Taj<sup>3</sup>,  
Taj Muhammad Hassani<sup>4</sup> and Wei Yunlin<sup>1\*</sup>**

\*Corresponding Author: **Wei Yunlin**  [weiyunlin18@gmail.com](mailto:weiyunlin18@gmail.com)

*Escherichia coli*, the enteric bacterium, is an ever-present constituent of all human beings and form a fraction of the normal flora of gut. It is debatably the most finely understood and extensively studied free-living organism on our planet. *Escherichia coli* is well acknowledged by the shortened name of *E. coli*. This article has introduced *E. coli* as a model organism in the biological study and also its several applications which have spawned from research of *E. coli*. *E. coli* is the first choice for researchers to investigate numerous basic biological processes which are essential for life and is the most extensively used organism in molecular genetics. The reason of widespread use of *E. coli* for study purpose is the ease of its maintenance and breeding in a laboratory environment plus its meticulous experimental advantages. As compared to other living organisms more is known about *E. coli* because of its simple nutritional requirements, rapid growth rate and most important it's well established genetics. Rate of cell division of *E. coli* is average of once in every 30 min, thus enabling quick environmental adaptation. This fast division rate of *E. coli* has helped in evolutionary experiments which are conducted in the laboratories.

**Keywords:** *Escherichia coli*, Model, Organism, Research, Field

## INTRODUCTION

*Escherichia coli* is a Gram-negative, non sporulating and facultative anaerobic rod. It is about 2.0 micrometers ( $\mu\text{m}$ ) in length and its diameter is 0.25-1.0  $\mu\text{m}$  as shown in Figure 1 (Kubitschek, 1990). Those strains which have flagella are motile. Structurally flagella have peritrichous arrangement (Darnton *et al.*, 2007). 37°C (98.6°F) is the optimal temperature for

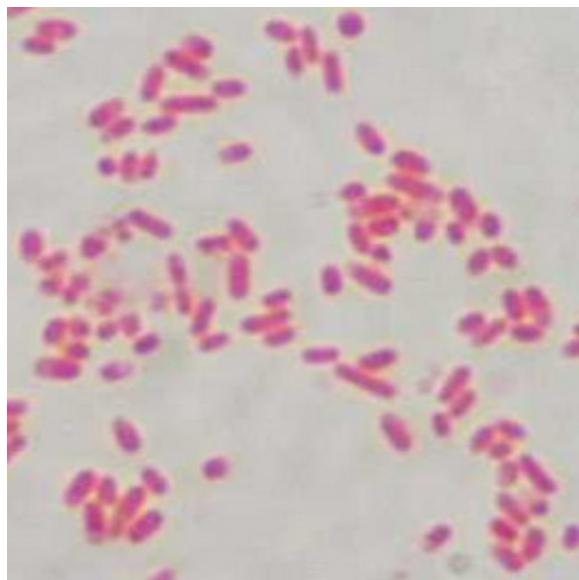
multiplication of *E. coli* but few of the laboratory strains can grow up to 49°C (120°F) of temperature (Fotadar, 2005). Multiplication can be driven by utilizing a large number of redox pairs involving 'reduction' of substrates like oxygen, fumarate, trimethylamine N-oxide and dimethyl sulfoxide plus 'oxidation' of substances like formic acid, pyruvic acid, amino acid and hydrogen (Ingledeew and Poole, 1984).

<sup>1</sup> Kunming University of Science and Technology, Yunnan China.

<sup>2</sup> Bolan Medical Hospital, Quetta, Balochistan, Pakistan.

<sup>3</sup> Centre for Advanced Studies in Vaccinology and Biotechnology, University of Balochistan, Balochistan, Pakistan.

<sup>4</sup> Food and Agriculture Organization, Balochistan, Pakistan.

**Figure 1: *Escherichia coli***

## LIFE CYCLE OF *ESCHERICHIA COLI*

In life cycle of *Escherichia coli*, there is division of one cell into two daughter cells. This process is known as binary fission (Figure 2). Under circumstances when no mutation occurs, the

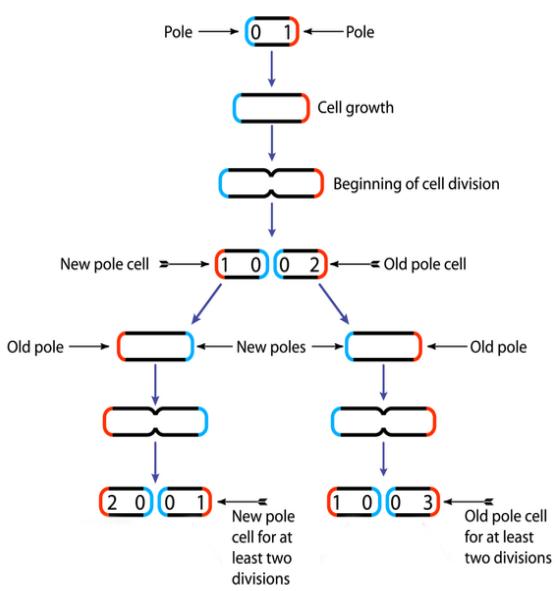
daughter cells are identical genetically to the parent cell. Thus there is “local doubling” of the *E. coli* population. However, it is not necessary that both daughter cells survive but *E. coli* population undergoes exponential growth, if the numbers of surviving daughter cells exceed unity on an average (Zwietering et al., 1990).

## ***ESCHERICHIA COLI TRANSMISSION***

*Escherichia coli* is found commonly in the distal part of intestine in warm-blooded organisms (endotherms) (Vogt and Dippold, 2005). They constitute about 0.1% of normal flora of gut (Eckburg et al., 2005). Major route of transmission for *E. coli* is oro-fecal after which bacterial pathogenic strains cause disease. *E. coli* cells can only survive outside the body for a limited period of time so they can be considered as ideal indicator organisms in order to test samples from environment for fecal contamination (Thompson and Andrea, 2007). However, research work carried out in this regard has showed that environmental samples may have *E. coli* strains that can survive for relatively long period of time even outside the host (Ishii and Sadowsky, 2008).

## ***ESCHERICHIA GENERA HISTORY***

A German pediatrician namedodor *Escherichia* in 1885 discovered *E. coli* in the feces of healthy individuals and named it *Bacterium coli* because of the fact it is found in the colon. Early prokaryotic classification placed *Bacterium coli* in a genera based on their motility and shape. Afterwards Ernst Haeckel's bacterial classification placed bacteria in the Monera kingdom (Escherich, 1885). In 1895 Migula reclassified

**Figure 2: Life Cycle of *Escherichia coli***

bacteria in the genus *Escherichia* which was named so after its discoverer (Castellani, 1919). This genus belongs to the bacterial group formally called "coliforms" which are member of the "the enterics" known as Enterobacteriaceae family (George and Garrity, 2005).

## **ESCHERICHIA COLI PHENOTYPIC DIVERSITY**

*Escherichia coli* includes a vast population of bacteria that demonstrate a very high degree of both phenotypic and genetic diversity. Taxonomic reclassification is required after looking at the genomic sequencing of a great number of isolates of the *E. coli* and also related bacteria (Krieg and Holt, 1984). *E. coli* is still one of the most varied bacterial specie and only about 20% of the genome seems to be common in all strains (Lukjancenko et al., 2010).

As a matter of fact, the members of *Shigella* genus (*S. flexneri*, *S. dysenteriae*, *S. sonnei* and *S. boydii*) must be classified as strains of *E. coli* from the evolutionary point of view. This phenomenon is termed as taxa in disguise. In the same way, other *E. coli* strains (e.g., the K-12 strain commonly used in recombinant DNA work) are very different, thus they warrant reclassification (Lan and Reeves, 2002).

A specie subgroup, which has distinctive characteristics thus distinguishing it from other, is called a strain. One can find these minute characteristic differences only at the molecular level which are responsible for changes in the lifecycle or physiology of the bacterium as for example, a strain may attain the ability to use a unique carbon source, take upon particular ecological niche, resist antimicrobial agents or gain pathogenic capacity. However, strains of *E. coli* are usually host-specific thus

making it easy to determine the source of fecal contamination in samples obtained from environment. A good example is that if researchers know that which *E. coli* strains are present in a sample of water, it will allow them to hypothesize that whether the contamination has its origin from a human or some other mammal and even from a bird (Thompson and Andrea, 2007).

On the bases of evolutionary relatedness, there is a common subdivision system of *E. coli* known as serotype. Serotype is based on antigens of surface (i.e., O-antigen which is a part of lipopolysaccharide layer, K-antigen, H: flagellin and capsule for example O157:H7) (Orskov et al., 1977). However it is common to quote only the serogroup that is the O-antigen. Presently upto 190 serogroups are known till date. The common strain of laboratory is non type able because it has a mutation which prevents the formation of an O-antigen (Stenutz et al., 2006).

New strains of *E. coli* evolved like all life forms, i.e., through the natural biological processes of horizontal gene transfer, gene duplication and mutation. About 18% laboratory strain MG1655 genome was acquired horizontally from *Salmonella* divergence (Lawrence and Ochman, 1998). All *E. coli* strains are derived from either *E. coli* B or *E. coli* K-12 strains. In microbiology, few strains developed traits which can be harmful to a animal host (Nataro and Kaper, 1998). O157:H7 is a more virulent strain which causes serious illness and even death in immunocompromised persons, the elderly people and the very young ( Hudault et al., 2001).

## **ESCHERICHIA COLI STRAINS**

Many of the *E. coli* strains have been

characterized and isolated. Most of the strains of *E. coli* which are commonly used in the research work are derived from Clifton's K-12 strain ( $\lambda^+$  F<sup>+</sup>; O16) and to a lesser extent from *Bacillus coli* strain (B strain; O7) (Brzuszkiewicz et al., 2011).

## DNA SEQUENCE OF *ESCHERICHIA COLI*

About 60 genomic sequences of *Shigella* and *Escherichia* species are available which have complete genomic sequences. A significant amount of diversity is seen when these sequences are compared. Over 20% of each genome represents those sequences which are present in each isolates and about 80% of genome can differ among the isolates (Lukjancenko et al., 2010). An individual genome consists about 4,000 to 5,500 genes, however there are more than 16,000 gene sequences in *E. coli* strains (the pan-genome). This fact reveals that a large variety of component genes exist which have been interpreted and about two-thirds of pan-genome of the *E. coli* originated in other species by horizontal gene transfer process (Zhaxybayeva and Doolittle, 2011).

## *ESCHERICHIA COLI* PROTEINS

Many *E. coli* proteins have been isolated and studied. A study purified 4,339 proteins from cultures of strain K-12 and found interacting partners for 2,667 proteins, many of which had unknown functions at that time (Arifuzzaman et al., 2006). But another study found 5,993 interactions between proteins of the same *E. coli* strain though this data showed little overlap with that of the above data (Hu et al., 2009).

## ***ESCHERICHIA COLI* AS A MODEL ORGANISM**

In a laboratory setting, the *E. coli* can be grown inexpensively and easily. *E. coli* has been widely studied for about 60 years. It is the most extensively investigated prokaryotic model organism and considered to be very important species in biotechnology and microbiology (Fang et al., 2002).

## **NONPATHOGENIC *ESCHERICHIA COLI* ADVANTAGES**

Nonpathogenic strains of *Escherichia coli* serve as probiotic agents in the field of medicine especially to treat various diseases of gastrointestinal tract (Grozdanov et al., 2004) which include inflammatory bowel disease (Kamada et al., 2005).

## **ROLE OF *ESCHERICHIA COLI* IN RESEARCH FIELD**

*E. coli* holds an important position in industrial microbiology and modern biological engineering because of its easy manipulation and also long history of its laboratory cultures (Lee, 1996). The research work of Herbert Boyer and Stanley Norman Cohen regarding use of restriction enzymes and plasmids in order to create recombinant DNA by *E. coli* became the base of biotechnology (Russo, 2003).

*E. coli* is considered to be a very flexible host for the heterologous proteins production (Cornelis, 2000). Recombinant protein production involves various protein expressions in *E. coli*. Plasmids have been used to introduce genes into the microbes by researchers which have lead to high level of protein expression. Such proteins can be produced by the fermentation process in

the industries at mass level. A very useful and important application of recombinant DNA technology was production of human insulin by *E. coli* manipulation (Tof and Ilanit, 1994).

Many folded forms of proteins have been successful in expressing them in *E. coli* which was previously thought to be difficult and even impossible. A good example is that proteins which have multiple disulphide bonds may have the ability to be produced in periplasmic space or in cytoplasm of those mutants having sufficiently oxidizing agent in order to allow the formation of disulphide-bonds (Bessette et al., 1999). On the other hand proteins that need post-translational modification glycosylation for function or stability use the system of N-linked glycosylation which is found in *Campylobacter jejuni* engineered into *E. coli* for their expression (Huang et al., 2012).

*E. coli* cells in modified form have been used in the development of vaccine, bioremediation, biofuels production (Summers and Rebecca, 2013) and formation of immobilized enzymes (Nic, 2013).

In microbiology studies, *E. coli* is widely used as a model organism. Unlike wild type strains, cultivated strains (e.g., *E. coli* K12) used in the laboratories are well-adapted to the lab environment and cannot survive in intestine. Many laboratory strains have lost their ability of biofilms formation. By this way wild type strains are protected from antibodies and also other chemical attacks, however, this requires expenditure of large number of material resources and energy (Fux et al., 2005).

*E. coli* was used as a model bacterium in order to describe bacterial conjugation (Lederberg et al., 1946) and it is still the primary model to study

this process. *E. coli* has helped in understanding phage genetics (Benzer, 1961).

## CONCLUSION

*Escherichia coli* is a natural mammalian gut bacteria used as a model organism for scientific research. *E. coli* is a single-celled organism that can be manipulated and killed with no ethical concerns. It has a rapid growth rate and is very easy to culture in laboratory. The *E. coli* can survive in uneven growth conditions including uneven temperatures, oxygen content, and nutrient availability. Most strains of *E. coli* are harmless, posing no threat to the scientists that use them. Finally, and most importantly, *E. coli* genetics are well-studied and can be manipulated easily. Genetic engineering using tools like conjugation and transduction allow scientists to genetically engineer *E. coli* strains to better understand genetic processes and test new ideas.

## ACKNOWLEDGMENT

I would like to express my very great appreciation to professor Ban Bo and Wang Nan, for their valuable and constructive suggestions during the planning and development of this review. My thanks and appreciations also go to my colleague in developing the article and people who have willingly helped me out with their abilities.

## REFERENCES

- Arifuzzaman M, Maeda M, Itoh A, Nishikata K, Takita C, Saito R, Ara T, Nakahigashi K, Huang H C, Hirai A, Tsuzuki K, Nakamura S, Altaf-Ul-Amin M, Oshima T, Baba T, Yamamoto N, Kawamura T, Ioka-Nakamichi T, Kitagawa M, Tomita M, Kanaya S, Wada C and Mori H (2006). "Large-scale

- identification of protein-protein interaction of *Escherichia coli* K-12", *Genome Res.*, Vol. 16, No. 5, pp. 686-91.
2. Benzer S (1961), "On the topography of the genetic fine structure", *Proc Natl Acad Sci.*, Vol. 47, No. 3, pp. 403-415.
  3. Bessette P H, Aslund F, Beckwith J and Georgiou G (1999), "Efficient folding of proteins with multiple disulfide bonds in the *Escherichia coli* cytoplasm", *Proc Natl Acad Sci.*, Vol. 96, No. 24, pp. 13703-13708.
  4. Brzuszkiewicz E, Thurmer A, Schuldes J, Leimbach A, Liesegang H, Meyer F D, Boelter J, Petersen H, Gottschalk G and Daniel R (2011), "Genome sequence analyses of two isolates from the recent *Escherichia coli* outbreak in Germany reveal the emergence of a new pathotype: Enterohemorrhagic *Escherichia coli* (EAHEC)", *Arch Microbiol.*, Vol. 193, No. 12, pp. 883-891.
  5. Castellani A and Chalmers A J (1919), "Manual of Tropical Medicine", 3<sup>rd</sup> Edition, Williams Wood and Co., New York.
  6. Cornelis P (2000), "Expressing genes in different *Escherichia coli* compartments", *Curr Opin Biotechnol.*, Vol. 11, No. 5, pp. 450-454.
  7. Darnton N C, Turner L, Rojevsky S and Berg H C (2007), "On torque and tumbling in swimming *Escherichia coli*", *J Bacteriol.*, Vol. 189, No. 5, pp. 1756-1764.
  8. Eckburg P B, Bik E M, Bernstein C N, Purdom E, Dethlefsen L, Sargent M, Gill S R, Nelson K E and Relman D A (2005), "Diversity of the human intestinal microbial flora", *Science.*, Vol. 308, No. 5728, pp. 1635-1638.
  9. Escherich T (1885), "Die Darmbakterien des Neugeborenen und Sauglings", *Fortschr Med.*, Vol. 3, pp. 515-522.
  10. Feng P, Weagant S and Grant M (2002), "Enumeration of *Escherichia coli* and the Coliform Bacteria", *Bacteriological Analytical Manual*, 8<sup>th</sup> Edition, FDA/Center for Food Safety & Applied Nutrition.
  11. Fotadar U, Zaveloff P and Terracio L (2005), "Growth of *Escherichia coli* at elevated temperatures", *J Basic Microbiol.*, Vol. 45, No. 5, pp. 403-404.
  12. Fux C A, Shirtliff M, Stoodley P and Costerton J W (2005), "Can laboratory reference strains mirror "real-world" pathogenesis", *Trends Microbiol.*, Vol. 13, No. 2, pp. 58-63.
  13. George M and Garrity (2005), "The Gammaproteobacteria", *Bergey's Manual of Systematic Bacteriology*, 2B (2<sup>nd</sup> ed.), New York, Springer, pp. 1108, ISBN 978-0-387-24144-9.
  14. Grozdanov L, Raasch C, Schulze J, Sonnenborn U, Gottschalk G, Hacker J and Dobrindt U (2004), "Analysis of the genome structure of the nonpathogenic probiotic *Escherichia coli* strain Nissle 1917", *J Bacteriol.*, Vol. 186, No. 16, pp. 5432-5441.
  15. Hu P, Janga S C, Babu M, Diaz-Mejia J J, Butland G, Yang W, Pogoutse O, Guo X, Phanse S, Wong P, Chandran S, Christopoulos C, Nazarians-Armavil A, Nasser N K, Musso G, Ali M, Nazemof N, Eroukova V, Golshani A, Paccanaro A, Greenblatt J F, Moreno-Hagelsieb G and

- Emili A (2009), "Global functional atlas of *Escherichia coli* encompassing previously uncharacterized proteins", *PLoS Biol.*, Vol. 7, No. 4, p. 96.
16. Huang C J, Lin H and Yang X (2012), "Industrial production of recombinant therapeutics in *Escherichia coli* and its recent advancements", *J Ind Microbiol Biotechnol.*, Vol. 39, No. 3, pp. 383-399.
17. Hudault S, Guignot J and Servin A L (2001), "*Escherichia coli* strains colonizing the gastrointestinal tract protect germ-free mice against *Salmonella typhimurium* infection", *Gut.*, Vol. 49, No. 1, pp. 47-55.
18. Ingledew W J and Poole R K (1984), "The respiratory chains of *Escherichia coli*", *Microbiol. Rev.*, Vol. 48, No. 3, pp. 222-271.
19. Ishii S and Sadowsky M J (2008), "*Escherichia coli* in the Environment: Implications for Water Quality and Human Health", *Microbes Environ.*, Vol. 23, No. 2, pp. 101-108.
20. Kamada N, Inoue N, Hisamatsu T, Okamoto S, Matsuoka K, Sato T, Chinen H, Hong K S, Yamada T, Suzuki Y, Suzuki T, Watanabe N, Tsuchimoto K and Hibi T (2005), "Nonpathogenic *Escherichia coli* strain Nissle1917 prevents murine acute and chronic colitis", *Inflamm Bowel Dis.*, Vol. 11, No. 5, pp. 455-463.
21. Krieg N R and Holt J G (1984), "*Bergey's Manual of Systematic Bacteriology*", (First ed.). Baltimore: The Williams & Wilkins Co. pp. 408-420, ISBN 0-683-04108-8.
22. Kubitschek H E (1990), "Cell volume increase in *Escherichia coli* after shifts to richer media", *J Bacteriol.*, Vol. 172, No. 1, pp. 94-101.
23. Lan R and Reeves P R (2002), "*Escherichia coli* in disguise: molecular origins of Shigella", *Microbes Infect.*, Vol. 4, No. 11, pp. 1125-1132.
24. Lawrence J G and Ochman H (1998), "Molecular archaeology of the *Escherichia coli* genome", *Proc Natl Acad Sci.*, Vol. 95, No. 16, pp. 9413-9417.
25. Lederberg J and Tatum E L (1946), "Gene recombination in *E. coli*", *Nature.*, Vol. 158, No. 4016, p. 558.
26. Lee S Y (1996), "High cell-density culture of *Escherichia coli*", *Trends Biotechnol.*, Vol. 14, No. 3, pp. 98-105.
27. Lukjancenko O, Wassenaar T M and Ussery D W (2010), "Comparison of 61 sequenced *Escherichia coli* genomes", *Microb Ecol.*, Vol. 60, No. 4, pp. 708-720.
28. Nataro J P and Kaper J B (1998), "Diarrheagenic *Escherichia coli*", *Clin Microbiol Rev.*, Vol. 11, No. 1, pp. 142-201.
29. Nic H (2013), "Bacteria-Powered Light Bulb Is Electricity-Free", <http://news.discovery.com>.
30. Orskov I, Orskov F, Jann B and Jann K (1977), "Serology, chemistry, and genetics of O and K antigens of *Escherichia coli*", *Bacteriol Rev.*, Vol. 41, No. 3, pp. 667-710.
31. Russo E (2003), "The birth of biotechnology", *Nature.*, Vol. 421, No. 6921, pp. 456-457.
32. Stenutz R, Weintraub A and Widmalm G (2006), "The structures of *Escherichia coli* O-polysaccharide antigens", *FEMS Microbiol Rev.*, Vol. 30, pp. 382-403.
33. Summers and Rebecca (2013), "Bacteria

- churn out first ever petrol-like biofuel New Scientist", [www.newscientist.com](http://www.newscientist.com).
34. Thompson and Andrea (2007), "E. coli Thrives in Beach Sands", *Live Science*.
  35. Tof and Ilanit (1994), "Recombinant DNA Technology in the Synthesis of Human Insulin", Little Tree Pty. Ltd.
  36. Vogt R L and Dippold L (2005), "Escherichia coli O157:H7 outbreak associated with consumption of ground beef", *Public Health Rep.*, Vol. 120, No. 2, pp. 174-178.
  37. Zhaxybayeva O and Doolittle W F (2011), "Lateral gene transfer", *Curr Biol.*, Vol. 21, No. 7, pp. R242– R246.
  38. Zwietering M H, Jongenburger I, Rombouts F M and Riet K V T (1990), "Modeling of the Bacterial Growth Curve", *Applied and environmental microbiology*, Vol. 56, No. 6, pp. 1875-1881.



**International Journal of Engineering Research and Science & Technology**  
**Hyderabad, INDIA. Ph: +91-09441351700, 09059645577**  
**E-mail: editorijerst@gmail.com or editor@ijerst.com**  
**Website: www.ijerst.com**

